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Evaluation of the sleep modulating effects of methanolic extracts of *Strychnos spinosa* and *Strychnos innocua* fruits in mice

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Copyright: C 2022 Abstract Sani et al. This is an This study aimed to evaluate the sleep modulating properties of Strychnos spinosa and Strychnos innocua fruits in Swiss albino mice. The depressant effect of the extracts on open-access article the central nervous system was studied using a diazepam-induced sleep model in mice. published under the terms of the Creative The mice were divided into four groups of six mice each. The test groups received Commons Attribution Strychnos spinosa and Strychnos innocua fruits extracts at the doses of 250, 500, and License which permits 1000 mg/kg body weight intraperitoneally. In contrast, the control group received distilled water at 10 ml/kg. Diazepam (Ranbaxy[®], India) was used as a standard drug and unrestricted use. distribution, administered 30 minutes after the initial treatment. The lethal doses (LD₅₀) of the and two extracts were estimated to be >5000 mg/kg. The results obtained from the reproduction in any medium, provided the onset of sleep of rats administered methanol extracts of Strychnos spinosa, and Strychnos innocua fruits showed that the extracts at 250, 500, and 1000 mg/kg, original author and source are credited. respectively, did not significantly (p >0.05) alter the onset of sleep when compared with the control. The onset of sleep in the groups treated with *Strychnos spinosa* fruit extracts indicated a dose-dependent decrease pattern. The study also revealed a prolonged duration of sleep in all the experimental groups treated with the two extracts compared to the control group. The treated groups of 250, 500 and 1000 mg/kg significantly (p < 0.05) prolonged the duration of diazepam-induced sleep in a Publication dose-dependent manner when compared to the control group. The group History: Received: 09-12-2021 treated with 1000 mg/kg of Strychnos innocua fruit extract was significant (p Revised: 20-02-2022 <0.05) compared to the control group. In conclusion, the high LD_{50} observed in Accepted: 25-03-2022 this study suggests that the two extracts are relatively safe and contain promising bioactive ingredients that cause sleep modulation in mice.

Keywords: Extract, Mice, Sleep modulation, Strychnos innocua, Strychnos spinosa

Introduction

Strychnos spinosa and S. innocua are flowering plants that belong to the family *Loganiaceae*. The genus Strychnos is widely distributed around tropical regions in the world. Strychnos possess indole alkaloids in their leaves, stems and roots, which have been reported to be poisonous. Strychnine and curare are the commonest alkaloids associated with the plant (Orwa et al., 2009). Strychnos is an important genus due to various reported therapeutic effect it possesses in Africa and Asia, such as therapy against fever, wound, snakebites, poisoning, tonic and emetic properties against gastrointestinal diseases, treatment against parasitic and skin diseases including leprosy. S. innocua is used in different regions of Africa to treat chronic malaria (Neuwinger, 1996). Scientists over the years have discovered alkaloids from plant-based sources are beneficial in the field of surgery. They are used as local and general anaesthetics to suppress brain activity, nerve transmission, muscle relaxation and control of pain (Brown et al., 2018). Various parts of Strychnos plant used for medicinal purposes include: decoction of the root to treat gonorrhea and chlamydia infection, fresh roots are used to treat snakebite and root infusions in Uganda to treat theileriosis in cattle (Tabuti et al., 2003; Orwa et al., 2009; Chinsembu, 2016). The bark and twigs are used to dilate the cervix during childbirth (Orwa et al., 2009), the leaves are used to treat malaria and also serve as antibacterial agent (Koné et al., 2004; Asase et al., 2005). The seeds have emetic properties and the fruit pulp is used as a remedy for dysentery and as eardrops (Orwa et al., 2009).

Toxicity of *Strychnos* species differ according to the part of the plant and this is associated with strychnine. The root is where strychnine undergo biosynthesis and it is rich in alkaloids and have a greater pharmacological activity than other plant parts. In India, pastoralist allow their cows to graze on the leaves of *Strychnos nux vomica* (Bisset, 1974). There is paucity of information regarding the modulatory effect of *Strychnos spp.* on sleep. Thus, this study was carried out to evaluate the sedative properties of *Strychnos spinosa* and *Strychnos innocua* fruits methanol extracts in mice.

Materials and Methods

Ethical clearance

The approval for the conduct of this study was obtained from Ahmadu Bello University Committee on Animal Use and Care (ABUCAUC) with approval number ABUCAUC/2018/077.

Experimental animal and management

A total of 72 Albino mice consisting of both sexes each weighing between 17 and 30 g were used for this study. They were fed with commercial poultry feed (Vitafeed[®]), while water was provided *ad libitum* throughout the study period in the Animal House, Department of Pharmaceutical Sciences, Ahmadu Bello University Zaria.

Experimental design

Plant collection and identification: The matured fruits of *Strychnos spinosa* and *Strychnos innocua* were randomly collected from different locations in Zaria, Kaduna State, Nigeria. The fruits were authenticated at the Herbarium, Department of Botany, Faculty of Life Sciences, Ahmadu Bello University Zaria, Nigeria and the plants were given a Voucher identification number of 901 and 902 for *Strychnos spinosa* and *Strychnos innocua* respectively. The fruits were stored at the Herbarium for reference purpose.

Methanol extraction of Strychnos spinosa and Strychnos innocua fruits: Methanol extraction was carried out at the Department of Pharmacognosy and Drug Development, Ahmadu Bello University Zaria, Nigeria. The method used was as described by Donatus & Ephraim (2009). The two fruits were dried in the shade and were pounded together using mortar and pestle to a coarse powder. The powder was packed into white cotton bags and put into Soxhlet apparatus (Sigma) and extracted exclusively using 3.5 L of 70% methanol (Sigma- Aldrich[®]) at 70°C.The methanolic extract was concentrated by evaporation at room temperature on warm waterbath (HH-S 21.6, double row six holes, HINOTEK China) to separate the solvent from the extract (oily liquid) at a temperature of 93.3°C.

Acute toxicity studies of *Strychnos spinosa* and *Strychnos innocua* fruit extracts: Twenty-four (24) mice were used for the intraperitoneal median lethal doses of the two extracts (12 each) using the method described by Lorke (1983).

Diazepam-induced sleep in mice: The method described by Rakotonirina *et al.* (2001) was used for the diazepam-induced sleep model for each of the extracts (consisting of 24 mice per extract). Each extract has 4 groups I-IV consist of 6 mice each and were treated with distilled water (10 mL/kg), 250, 500 and 1,000 mg/kg of the fruit extract (*S. spinosa/S. innocua*) respectively, intraperiotoneally. Thirty minutes later, diazepam (Roche[®]) at 2mg/kg intraperitoneally was administered to all the groups

respectively. All the animals for the two fruit extracts were observed continuously for the induction time, time of onset of action and recovery time, until the mice were fully recovered.

Data analysis

Values obtained were expressed as mean \pm standard error of mean (SEM). One-way analysis of variance (ANOVA) was used followed by Tukey's *post-hoc* test for multiple comparisons of groups. GraphPad Prism version 8.0.0 for windows (GraphPad software San Diego, California, USA) was used for the analysis. Values of p < 0.05 were considered significant.

Results

The median lethal dose of the two extracts was estimated to be greater than 5000 mg/kg suggesting that they are relatively safe intraperitoneally. No significant (p> 0.05) difference in the onset of sleep was observed in all the groups treated with *Strychnos spinosa* fruit when compared to the control. (Table 1). There was a significant (p <0.05) increase in the duration of sleep in *S. spinosa* extracts (80.10 \pm 0.24; 115.40 \pm 0.51 and 138.80 \pm 0.37) respectively in all the treated groups in a dose-depended manner when compared to the control (17.60 \pm 4.2) (Table 2). No significant (p> 0.05) difference in the onset of sleep

was observed in all the groups treated with *Strychnos innocua* fruit when compared to the control. (Table 3). The sleep modulatory effect of *Strychnos innocua* fruit extract on diazepam-induced sleep is shown in Table 4. The group treated with the highest dose (1000 mg/kg) showed significant (p <0.05) increase (79.40 ±1.76) in the duration of sleep when compared to the control (17.00 ± 4.23) (Table 4).

Discussion

The lethal intraperitoneal doses (LD₅₀) of the two extracts were estimated to be greater than 5000 mg/kg, hence suggesting that the extracts are safe (Matsumura, 1985; Loomis & Hayes, 1996) or practically nontoxic (Lorke, 1983). The safety of these extracts observed in this study agrees with the earlier cytotoxicity study on S. spinosa by Adamu et al. (2014), who reported an LC50 of 361.48 µg/ml on monkey kidney Vero cells In vitro using the MTT (3-(4,5-dimethylythiazol-2-yl)-2,5diphenyl-2H-tetrazolium hydrobromide) assay. The high value of LD₅₀ established in this study has revealed the safe nature of these two extracts and may explain the reasons behind animals and humans eating the fruits without visible detrimental effects. The results obtained on the onset of sleep of animals treated with methanol

Table 1: Effect of methanol extract of Stry	<i>chnos spinosa</i> fruit on diazer	pam-induced sleep-in mice	(onset of sleep)

Treatment (mg/kg)	Onset of sleep (min)
Control (10 ml/kg) distilled water + Diazepam (2 mg/kg)	4.60 ± 0.40
Strychnos spinosa (250 mg/kg) + Diazepam (2 mg/kg)	4.20 ± 0.14
Strychnos spinosa (500 mg/kg) + Diazepam (2 mg/kg)	4.00 ± 0.31
Strychnos spinosa (1000 mg/kg) +Diazepam (2 mg/kg)	3.70 ± 0.27
Values are expressed as mean $\pm SEM(n-6)$	

Values are expressed as mean ± SEM (n=6)

Table 2: Effect of methanol extract of *Strychnos spinosa* fruit on diazepam-induced sleep-in mice (duration of sleep)

Treatment (mg/kg)	Duration of sleep (min)
Control (10 ml/kg) distilled water + Diazepam (2 mg/kg)	17.60 ± 4.20
Strychnos spinosa (250 mg/kg) + Diazepam (2 mg/kg)	80.10 ± 0.24*
Strychnos spinosa (500 mg/kg) + Diazepam (2 mg/kg)	115.40 ± 0.51**
Strychnos spinosa (1000 mg/kg) + Diazepam (2 mg/kg)	138. 80 ± 0.37***
	*** . 0.0004

Values are expressed as mean ± SEM (n=6), *p < 0.05, **p < 0.001, ***p < 0.0001 compared to control

Table 3: Effect of methanol extract of Strychnos innocua fruit on diazepam-induced sle	leep-in mice (onset of
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sleep)		
Treatment (mg/kg)	Onset of sleep (min)	
Control (10 ml/kg) distilled water + Diazepam (2 mg/kg)	4.60 ± 0.40	
Strychnos innocua (250 mg/kg) + Diazepam (2 mg/kg)	4.4 ± 0.24	
Strychnos innocua (500 mg/kg) + Diazepam (2 mg/kg)	4.40 ± 0.51	
Strychnos innocua (1000 mg/kg) + Diazepam (2 mg/kg)	3.8 ± 0.37	

Values are expressed as mean ± SEM (n=6)

Treatment (mg/kg)	Duration of sleep (min)
Control (10 ml/kg) distilled water + Diazepam (2 mg/kg)	17.00 ± 4.23
Strychnos innocua (250 mg/kg) + Diazepam (2 mg/kg)	53.80 ± 1.72*
Strychnos innocua (500 mg/kg) + Diazepam (2 mg/kg)	51.80 ± 1.44*
Strychnos innocua (1000 mg/kg) + Diazepam (2 mg/kg)	79.40 ± 1.76**

Table 4: Effect of methanol extract of Strychnos innocua fruit on diazepam-induced sleep-in mice (duration of sleep)

Values are expressed as mean ± SEM (n=6), *p < 0.05, **p < 0.001 compared to control

extract of Strychnos spinosa and Strychnos innocua fruits indicated that the extract at the doses of 250-1000 mg/kg did not significantly alter the onset of sleep compared with the control. The onset of sleep in the groups treated with Strychnos spinosa fruit extract showed a dose dependent decrease pattern. The study also revealed a prolonged duration of sleep in all the experimental groups treated with the two extracts compared with the control group. The prolonged duration of sleep was also observed to be statistically significant only in the group treated with 1000 mg/kg compared to the control group. The faster onsets and the significantly prolonged duration of diazepam-induced sleep time observed in this study indicate that the two methanol extracts potentiate diazepam induced sleep in mice and thus possess a sleep-inducing property (Rakotonorina et al., 2001). The sedative properties of these two fruits extracts may be due to the presence of bio-active ingredients such as alkaloids, saponins, and flavonoids. Several researchers have reported that alkaloids, saponins, and flavonoids are responsible for sedation (Won et al., 1980; Wagner et al., 1983; Dubios et al., 1986; Amos et al., 2001; Musa et al., 2006; Viswanatha-Swamy et al., 2006; Musa et al., 2008). The results from this study demonstrate a relationship between the CNS depressant effect of the two extracts and diazepam, which could suggest that the extract of S. pinosa and S. inocua mimics the action of benzodiazepines.

Several works conducted on phytochemical investigation suggest that flavonoids and neuroactive steroids are ligands for GABA_A receptors in the CNS, indicating that they can act as benzodiazepine-like agents (Protapaditya *et al.*, 2011). It has also been reported that some flavonoids exhibit high-affinity binding to the benzodiazepine site of GABA_A receptors. Saponins and Triterpenoids have also been agonistic activities at the GABA_A receptor complex (Uma *et al.*, 2011; Kumaresan *et al.*, 2011; Khatoon *et al.*, 2014), suggesting that they act as benzodiazepines-like molecules. Therefore, the CNS depressant activity exhibited by these extracts may

be due to the phytoconstituents present in them. Even though there is no evidence on the specific constituent responsible for the CNS depressant effects.

We, therefore, predict that the action of these extracts may be due to the direct activation of GABA receptors (Kavita *et al.*, 2013). It may also be due to enhanced affinity for GABA or an increase in the duration of the GABA-gated channel opening (Shans-Ud-Doha *et al.*, 2013) or by potentiating GABAergic inhibition in the CNS via membrane hyperpolarisation leading to a reduction in the firing rate of critical neurons in the brain (Kavita *et al.*, 2013)

Conclusively, the relatively low toxicity and the Central Nervous Depression effects of these extracts, as demonstrated in this study, suggest that the two extracts are relatively safe and contain bioactive ingredients that cause sleep modulation in mice.

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Conflict of interest

The authors declare that there is no conflict of interest.

References

- Adamu II, Maurice DA, Jean PD, Mohammed A, Rabiu AM, Joseph OA & Jacobus NE. (2014). Some Strychnos spinosa (Loganiaceae) leaf and fractions extracts have good antimicrobial activities and low cytotoxicities. BMC Complementary and Alternative Medicine, doi.10.1186/1472-6882-14-456.
- Amos S, Kolawole E, Wambebe C & Gamaniel K (2001). Behavioral effects of aqueous extract of

Guieresenegalensis in mice and rats. *Phytomedical*, **8**(5): 356-361.

- Asase A, Oteng-Yeboah AA, Odamtten GT & Simmonds MSJ (2005). Ethnobotanical study of some Ghanaian anti-malarial plants. Journal of *Ethnopharmacology*, **99**(2): 273–279.
- Bisset NG (1974). The Asian species of Strychnos. Part III. The ethnobotany. *Lloydia*, **37**(1): 62–107.
- Brown EN, Pavone KJ & Naranjo M (2018). Multimodal general anesthesia: Theory and practice. *Anesthesia and Analgesia*,**127**(5): 1246-1258.
- Chinsembu KC (2016). Ethnobotanical study of medicinal flora utilised by traditional healers in the management of sexually transmitted infections in Sesheke District, Western Province, Zambia. Revista Brasileira de Farmacognosia. Brazillian Journal of Pharmacognosy, **26**(2): 268–274.
- Donatus EO & Ephraim CI (2009). Isolation, Characterisation and Antibacterial activity of alkaloid from *Datura metel*Linn leaves. *African Journal of Pharmacology*, **3**(5): 277-281.
- Dubios MA, Hyas M & Wagnar H (1986). Cussonosides A and B two triterpenes saponins from *Casseniabateri. PLantammedica*, **56**(2): 80-83.
- Kavita G, Vijay KL & Shivesh J (2013). Anticonvulsant potential of ethanol extracts and their solvent partitioned fractions from Flemingiastrobilifera root. *Pharmacognosy Research*, **5**(4): 265–270.
- Khatoon MM, Khatun MH, Islam ME & Parvin MS (2014). Analgesic, antibacterial and central nervous system depressant activities of Albiziaproceraleaves. *Asian Pacific Journal of Tropical Biomedicine*, **4**(4): 279–284.
- Koné WM, Atindehou KK, Terreaux C, Hostettmann C, Traoré D & Dosso M (2004). Traditional medicine in North C^ote-d'Ivoire: screening of 50 medicinal plants for antibacterial activity. *Journal of Ethnopharmacology*, **93**(1): 43-49.
- Kumaresan PT, Asish T & Vijaya C (2011). Neuropharmacological activity of Lippianodiflora Linn. Pharmacognosy Research, 3(3): 94–200.
- Loomis TA & Hayes AW (1996). Loomis Essentials of Toxicology, fourth edition, California, Academic press, Pp 56-60.

- Lorke D (1983). A new approach to practical acute toxicity testing. *Archives of Toxicology*, **54**(4): 275-287.
- Matsumura F (1985). Toxicology of pesticides, Plenum Press, London, Pp 24-26.
- Musa AM, Yaro AH & Danjuma NM (2006). Preliminary phytochemical screening and Central Nervous Depressant activity of the stem bark of *Ficustoningi* Blums. *Biology Environmental Science and Tropical Journal*, **3**(2): 1-6.
- Musa AM, Yaro AH, Usman H, Magaji MG & Habu M (2008). Phytochemical and some neuropharmacological studies on the methanol leaf extract of *Cissuscornifolia* (Vitaceae) in mice. *International Journal Pharmacology*, **4**(2): 145-148.
- Neuwinger HD (1996). African ethnobotany poisons and drugs: Chemistry, Pharmacology, Toxicology. Chapman & Hall, London. Pp 941.
- Orwa C, Mutua A, Kindt R, Jamnadass R & Anthony S (2009). Agroforestree Database: A tree Reference and Selection Guide Version 4.0. World Agroforestry Centre, Kenya. <u>http://www.worldagroforestry.org/sites/tre</u> <u>edbs/treedatabases.asp</u>, retrieved 05-12-2021.
- Protapaditya D, Sangita C, Priyanka C & Sanjib B (2011). Neuropharmacological properties of MikaniaScandens (L.) Willd. (Asteraceae). Journal of Advanced Pharmaceutical Technology and Research, **2**(4): 255–259.
- Rakotonirina VS, EN Bum A, Rakotonirina & M Bopelet (2001). Sedative properties of the decoctionof the rhizome of Cyperusarticularis.*Fitoterapia*, **72**(1): 22-29.
- Shans-Ud-Doha KM, Zobaer AM, Sitesh CB & Nazmul Q (2013). Antinociceptive, antiinflammatory, antimicrobialand central nervous system depressant activities of ethanolic extract of leaves and roots of Gomphostemmaparviflorum var. parviflorum wall. *Pharmacognosy Research*, **5**(4): 233–240.
- Tabuti JRS, Dhilliona SS & Lye KA (2003). Ethnoveterinary medicines for cattle (*Bosindicus*) in Bulamogi County, Uganda: Plant species and mode of use. *Journal of Ethnopharmacology*, **88**(2-3): 279-286.
- Uma AB, Radha Y, Prachi DP, Mandar RZ & Rahul SS (2011). Study of central nervous system depressant and behavioral activity of an ethanol extract of Achyranthesaspera

(Agadha) in different animal models. International Journal of Applied and Basic Medical Research. **1**(2): 104–108.

- Viswanatha-Swamy AHM, Thippeswami AHM, Manjala DV & MeahendraKumar CB (2006). Some neuropharmacological effects of the methanol root extract of *Cissusquadrangularis* in mice. *African Journal of Biomedical Resource*, **9**(2): 69-75.
- Wagner H, Ott S, Jureie K, Morton J & Neszmelyi A (1983). Chemistry 13C NMR study and pharmacology of two saponins from *Colubrinaasiatica. Planta Medica*, **48**(7): 136-141.
- Won SW, Kuk HS & Sam SK (1980). Chemistry and pharmacology of flavones-C glycosides from *Zizphus* seeds. *Korean Journal of Pharmacology*, **11**(3-4): 141-148.