

Esonu et al. /Sokoto Journal of Veterinary Sciences, **18**(1): 13 - 17.

Prevalence of *Salmonella* organisms in fresh and smoke-dried fish within parts of Kaduna metropolis, Kaduna State, Nigeria

DO Esonu*, BV Maikai & AJ Oghumu

Department of Veterinary Public Health and Preventive Medicine, Faculty of Veterinary Medicine, Ahmadu Bello University Zaria, Kaduna State Nigeria

*Correspondence: Tel.: +234 8061604710; E-mail: esonu25@gmail.com

Copyright: © 2020	Abstract
Esonu <i>et al</i> . This is an	Aquatic environments are the major reservoirs of Salmonella and fishery products
open-access article	have been recognized as major carrier of food-borne pathogens. To determine the
published under the	prevalence of Salmonella in fish, 112 fresh and smoke-dried fish samples were pre-
terms of the Creative	enriched and enriched with buffered peptone water and selenite broth respectively,
Commons Attribution	before plating on Desoxycholate Citrate Agar. Of the 112 samples, 75% (84/112)
License which permits	were contaminated with non-lactose fermenters. Upon subjecting them to
unrestricted use,	biochemical tests, 13.1% (11/84) of the Salmonella suspects showed reactions
distribution, and	consistent with that of Salmonella species, Proteus 65.5% (55/84) and Citrobacter
reproduction in any	21.4% (18/84) species. The total prevalence of Salmonella out of the 112 fish
medium, provided the	sampled was 9.8% (11/112). Salmonella species was slightly higher in the fiber tank
original author and source are credited.	farm 12.9% (95% CI: 5.134-28.852) than in the earthen pond 12.1% (95% CI: 4.816-
source are created.	27.326) but this association was not significant (Fishers exact test=1.0). Prevalence
	was higher in fresh fish 12.5% (95% CI: 6.472-22.775) compared to smoke-dried fish
	6.25% (95% CI: 2.148-16.835), though this association was not statistically significant
	(Fishers exact test= 0.347). Among the dried fish, the prevalence was higher 9.5%
	(95% CI: 0.017-0.289) in the weight range 30-49.9g compared to the smallest weight
	range of 10-29.9g with prevalence of 4.0% (95% CI: 0.002-0.195). For the fresh fish,
	the prevalence was higher 14.04% (95% CI: 7.287-25.324) in shorter length fish of 20-
	39cm than the lengthier ones of 40-49cm with 0.0% (95% CI: 0.000-35.433)
	prevalence. This study has demonstrated the presence of Salmonella species in fresh
Publication History:	and smoke-dried fish in parts of Kaduna metropolis, Kaduna State, Nigeria. This is of
Received: 20-09-2019	public health significance and poses a potential risk especially among
Accepted: 25-01-2020	immunocompromised consumers.

Keywords: Biochemical test, Earthen pond, Fresh fish, Fiber tank, Prevalence, Salmonella, Smoke-dried fish

Introduction

Salmonella species are the leading causes of acute gastroenteritis in several countries (Soltan *et al.,* 2009). Salmonellosis is the most common food-borne disease in both developing and developed countries, although incidence rates vary between humans and

animals. *Salmonella* species are important sources of contamination of the environment and the food chain (Ponce *et al.*, 2008). *Salmonella* serovars are zoonotic, leading food borne pathogens, responsible for outbreaks of both human and animal diseases and

have important health significance worldwide. There are several transmission routes for salmonellosis, but the majority of human infection are derived from consumption of contaminated foods (Mehemet *et al.*, 2003) such as insufficiently cooked meat, fish or improperly pasteurized milk and milk products (Jackson *et al.*, 2007). *Salmonella* infections can lead to numerous clinical conditions, such as enteric (typhoid) fever, uncomplicated enterocolitis, and systemic infections by non-typhoid microorganisms (Bailey & Maurer, 2005).

Fish is a very important source of animal protein in the diets of man. They constitute about 60% of the total protein intake in adults especially in the rural areas (Akise et al., 2013). In Nigeria, fish is eaten fresh, preserved or processed (smoked) and form a much-cherished delicacy that cuts across socioeconomic, age, religious and educational barriers (Adebayo-Tayo et al., 2008). Fish that is sold to the general public is often subjected to several unhygienic processes. Water used in washing the fish can also serve as source of contamination. If the fish is not properly washed, they can serve as vehicles for transmission of pathogenic organisms (Bailey & Maurer, 2005). Therefore, there is need to evaluate the safety of fish sold for human consumption because consumption of either fresh or smoked fish has been associated with gastroenteritis which indicates lack of proper hygiene practices during fish handling and preservation. This can constitute a food safety hazard. It is therefore pertinent to determine the prevalence of this organism in fresh and smoked fish.

Bacterial culture, isolation and biochemical characterization identification method has been used for the laboratory diagnosis of *Salmonella* organisms since it has been proven to be reliable. These cultural methods for the detection of *Salmonella* are also relatively easy to perform, cheap and readily available hence the use in this research study.

Materials and Methods

Study area

This study was conducted in Kawo and Ungwan Shanu districts of Igabi and Kaduna North Local Government Areas of Kaduna State respectively. The geographical coordinates of Kawo are 10° 34' 44'' North, 7° 26' 56'' East while that of Ungwan Shanu are 10° 31' 47'' North and 7° 21' 41'' East (Google map, 2014).

Sample collection

A total of 112 fish samples comprising of 64 fresh and 48 smoke-dried fishes, were randomly collected between July and September, 2014 using systematic random sampling method. The fresh fish samples were collected from fish farms located in Ungwan Shanu district. Every 3rd fish picked from the net was used for the study. The farms are made up of earthen ponds and fiber tanks while the smoke-dried fish were bought from Kawo market in Kawo district. The fish samples were purchased exactly the way they were sold to other consumers. The samples were packed in sterile polythene bags and transported to the bacterial zoonosis laboratory in the Department of Veterinary Public Health and Preventive Medicine, Ahmadu Bello University, Zaria. The fish samples were kept in the refrigerator not longer than twelve (12) hours if they were not immediately processed. Before the processing of each sample, the total weight of each fish was determined and in addition, total length of the fresh fish was also determined.

Laboratory procedures

From each sample collected, 10 grams (comprising the intestine, gills and parts of the skin) were weighed out using weighing balance and cut into smaller pieces after which it was placed in a large sterile white polythene and 90mls of buffered peptone water was added to it. The samples were then homogenized in the 90mls peptone water for 5 minutes using a laboratory blender (Stomacher L-B 400). This gives a dilution factor of 1:10. The homogenized samples were then incubated at 37°C for 24hours. From the clear middle layer (below the upper fat layer) of the homogenate, 1ml was pipetted and inoculated into 5mls of selenite broth. This was incubated at 37°C for 24 hours.

Using a sterile Pasteur's loop, a loop full of incubated selenite broth was streaked on Desoxycholate Citrate Agar to ensure the growth of isolated colonies. The plates were labeled, incubated at 37°C for 24 hours. The colonial morphology on the plate was then appraised and those colonies that were colorless with black centers indicating non-lactose fermenters, were stored on Nutrient Agar slants in bijour bottles and incubated at 37°C for 24 hours and kept in the refrigerator at 4°C pending biochemical characterization.

The purity of the isolates stored on Nutrient Agar slants was ascertained by plating on Desoxycholate Citrate Agar before stabbing and streaking on the Triple Sugar Iron (TSI) and this was incubated at 37°C for 24 hours. Typical *Salmonella* suspects give alkaline and acids reaction on the slant and butt of the TSI respectively, with or without hydrogen sulphide gas production. Isolates were also streaked on prepared urea and citrate agar slants. These were also

incubated at 37°C for 24 hours. *Salmonella* suspects maintained the original color of urea and change the green color of citrate to blue, indicating urea negative and citrate positive respectively.

Furthermore, Sulfide Indole Motility (SIM) agar was also inoculated by stabbing to test for motility, sulphur and indole production. Presence of cloud around stab-line after incubating for 24 hours at 37°C indicates motile organism, also black color formation indicates production of hydrogen sulphide. About 0.5ml of Kovac's reagent was added to SIM tubes and shaken. A pinkish color indicates indole production. Lastly, methyl red voges proskeur (MR-VP) was also inoculated and incubated at 37°C for 24 hours and 2 drops of methyl red was added, pinkish coloration indicates methyl red positive.

Data analysis

Data was analyzed using Statistical Package for Social Science version 20.0. Fisher's exact test was used to test for association between prevalence of *Salmonella* in fresh and smoke-dried fish and factors such as weight, length, preservation and management practices of fish. $P \le 0.05$ was considered significant.

Results

On processing the one hundred and twelve (112) samples collected, 75% (84/112) yielded non-lactose fermenting colonies upon plating on Desoxycholate Citrate Agar and were considered *Salmonella* suspects. The suspects were then subjected to biochemical tests and 13.1% (11/84) of them showed reactions consistent with that of *Salmonella* species while the remaining 86.9% (73/84) that were not *Salmonella*, showed reactions typical for *Proteus* 65.5% (55/84) and *Citrobacter* 21.4% (18/84) (Table 1).

The total prevalence of *Salmonella* out of the 112 fish sampled was 9.8% (11/112). Information on management practices obtained from each of the farms showed that fiber tank farm has a better management practice than that of the earthen pond in terms of hygienic practices. Though *Salmonella* spp. was slightly higher in the fiber tank farm 12.9% (95% CI: 5.134-28.852) than in the earthen pond 12.1% (95% CI: 4.816-27.326) but there was no significant association (Fisher's exact test = 1.0) between *Salmonella* spp. and management practice. The isolation of *Salmonella* with respect to the preservation method of the fish showed that fresh fish was higher 12.5% (95% CI: 6.472-22.775) compared to smoke-dried fish 6.25% (95% CI: 2.148-

16.835), though there was no significant association (Fisher's exact test = 0.347) between Salmonella spp. and preservation method. Among the dried fish, the prevalence was higher 9.5% (95% CI: 0.017-0.289) in the weight range of 30 - 49.9g compared to the smallest weight range of 10 – 29.9g with 4.0% (95% CI: 0.002-0.195) prevalence. No Salmonella was seen in the bigger 50 – 69.9g fish. This is in contrast to the fresh fish category where all the Salmonella positive fresh fish 14.0% (95% CI: 7.287-25.324) were isolated in the smallest weight range of 150-499g. No Salmonella was isolated in the 500-799g and 800-1099g weight ranges. It was observed that the prevalence was higher 14.04% (95% CI: 7.287-25.324) in shorter length fish of 20-39cm than the lengthier ones of 40-49cm 0.0% (95% CI: 0.000-35.433) (Table 2).

Discussion

The result of this study shows the presence of Salmonella, Proteus and Citrobacter in fresh and smoke-dried fish obtained from Igabi and Kaduna North Local Government Areas of Kaduna State. The prevalence of Salmonella 9.8% (11/112) that was gotten out of the total number of fish sampled is of public health significance as no Salmonella is expected to be present in food meant for consumption. Though, Agu et al. (2013) reported a higher prevalence of 20% in Benin, Nigeria while Raufu et al. (2014) reported a prevalence of 11.5% in Catfish reared in Maiduguri, sub-Saharah, Nigeria. The isolation of *Salmonella* from fish in this study may either be introduced through handling post-harvest, or contamination via the water used. Also, environmental contamination may have contributed to the isolation of other organisms such as Proteus and Citrobacter. Likewise, materials used for the preparation or the holding time may have contributed to the contamination (Uzeh et al., 2006; Ogbonna et al., 2012).

The slightly higher rate in the fiber tank than the earthen pond may be attributed to contamination from other sources such as feed or utensils used other than those from water or environment probably because there were no strict biosecurity measures

Table 1: Prevalence of bacterial isolates in fresh andsmoke-dried fish in parts of Kaduna metropolis,Kaduna State.

itadama otator			
Pathogens	Number positive (%)		
Salmonella spp.	11 (13.1)		
Proteus spp.	55 (65.5)		
Citrobacter spp.	18(21.4)		
Total	84 (75.0)		

Factors			Number	Number	Fisher's
		exami	examined	positive	exact test
Preservation	Preservation	Fresh fish	64	8 (12.5)ª	0.347
Method		Smoke-dried fish	48	3 (6.25) ^b	
Management	Fresh fish	Fiber tank	31	4 (12.9) ^c	1.0
practice		Earthen pond	33	4 (12.1) ^d	
Weight (g)	Smoke-dried fish	10-29.9	25	1 (4.0) ^e	
		30-49.9	21	2 (9.5) ^f	
		50-69.9	2	0 (0.0) ^g	
	Fresh fish	150-499	57	8 (14.0) ^h	
		500-799	6	0 (0.0) ⁱ	
		800-1099	1	0 (0.0) ^j	
Length (cm)	Fresh fish	20-39	57	8 (14.04) ^k	
,		40-49	7	0 (0.0)	
Total			112	11 (9.8) ^m	

Table 2: Prevalence of Salmonella spp. in fresh and smoke-dried fish in parts of Kaduna metropolis, Kaduna state.

95% CI: a= 6.472-22.775; b= 2.148-16.835; c=5.134-28.852; d= 4.816-27.326; e= 0.002-0.195; f= 0.017-0.289; g=

0.000-0.822; h= 7.287-25.324; i= 0.000-39.033; j= 0.000-94.871; l= 7.287-25.324; m= 0.000-35.433

employed in the farm. The higher contamination in fresh fish compared to the smoke-dried ones may be attributed to prolonged heat treatment the dried fish was subjected to during the process of preservation which must have helped in reducing the risk of Salmonella contamination of the smoke-dried fish. In this study, bigger fish were observed to have lower or no Salmonella as compared to smaller ones. This may be because microbial contamination may cover more of the entire surface area of a smaller organism faster than a larger organism and since it was not the entire fish that was processed, one can likely miss out some contaminated part more easily in a bigger fish than in a smaller one. This also applies in the case of lengthier fish as compared to smaller ones. This could also be due to the sampling method as more small and shorter length fish were sampled. This finding seems to be in contrast to the report of Emere & Egbe (2006) and Omeji et al. (2011) on parasite eggs and oocysts in fish whereby it was more in bigger and lengthier ones compared to smaller ones.

This study has demonstrated the presence of *Salmonella* in fish. Though the prevalence was relatively low, it is still of public health significance. We therefore recommend proper cooking of fish at all times before consumption and improvements in the detection and investigation of foodborne illnesses so that possible source of infection can be traced to ensure proper control measures.

Acknowledgement

We sincerely appreciate the efforts of laboratory staff of the Department of Veterinary Public Health and

Preventive Medicine, Faculty of Veterinary Medicine, Ahmadu Bello University, Zaria, in seeing to the completion of this research work.

Conflicts of Interest

The authors declare no conflict of interest.

References

- Adebayo-Tayo BC, Onilude AA & Patrick UG (2008). Mycoflora of Smoke-Dried Fishes Sold in Uyo, Eastern Nigeria. *World Journal* of Agricultural Science, **4**(3): 346-350.
- Agu KC, Ukponmwan IO, Orji MU, Awah NS, Anaso CI & Udemezue OI (2013). Prevalence of pathogenic bacteria in smoked fish sold in major retail markets in Benin, Nigeria. International Journal of Applied Sciences and Engineering, **1**(1): 1-4.
- Akise OG, Abolagba OJ & Eyong MM. (2013). Mycoflora of three fish species smoke-dried using rubber wood (*Hevea brassillensis*) in Nigeria. *Greener Journal of Agricultural Sciences*, **3**(5): 396-402.
- Bailey JS & Maurer JJ (2005). *Salmonella* Species. In Food Microbiology: An Introduction (TJ Montville, KR Matthews, editors). ASM Press, Washington DC. Pp 85-99.
- Emere MC & Egbe NEL (2006). Protozoan parasites of Synodonits clarias (a fresh water fish in river Kaduna). Best Journal, **3** (3): 58-64.
- Google map (2014). Google Maps. http://maps.google.com/.

Jackson CR, Cray PJ, Haro JH & Mcglinchey B (2007). Prevalence of *Salmonella* in beef and dairy cattle and potential pathogenicity of their isolates.

www.Msda.gov/research/publications/htm, retrieved 12-12-2013.

- Mehemet C, John S, Kendendail AP & Gary CS (2003). Effect of acid adaptation on the inactivation of *Salmonella* during storage of beef jerky treated with maimaid. *International Journal* of Food Microbiology, **89**(1): 51-65.
- Ogbonna IO, Danladi MS, Akinwusire O & Odu CE (2012). Microbiological safety and proximate composition of suya street store at ambient temperature for six hours from Maiduguri; Northern Nigeria. *Internet Journal of Food Safety*, **14**(1): 11-16.
- Omeji S, Solomon SG & Idoga ES (2011). A Comparative study of the common protozoan parasites of *Clarias gariepinus* from the wild and cultured environments in

Benue state, Nigeria. *Journal of Parasitology Research*, doi: 10.1155/2011/916489.

- Ponce E, Khan AA, Cheng CM, Summage-West C & Cerniglia CE (2008). Prevalence and characterization of *Salmonella enterica* serovar Weltevreden from seafood. *Journal* of Food Microbiology, **25**(1): 29-35.
- Raufu IA, Lawan FA, Bello HS, Musa AS, Ameh JA & Ambali AG (2014). Occurrence and antimicrobial susceptibility profiles of *Salmonella* serovars from fish in Maiduguri, sub-Saharah, Nigeria. *The Egyptian Journal* of Aquatic Research, **40**(1): 59-63.
- Soltan MM, Taremi L, Gachkar S, Moderresi M & Sanaei R (2009). Characterization of antibiotic resistant patterns of *Salmonella* serotypes isolated from beef and chicken samples in Tehran. Jundishapur. *Journal of Microbiology*, **2**(4): 124-131.
- Uzeh RE, Ohenhen RE & Adeniji OO (2006). Bacterial contamination of Tsire-Suya, Nigerian Meat Product. *Journal of Nutrition*, **5**(5): 458-460.