SHORT COMMUNICATION



Sokoto Journal of Veterinary Sciences

(P-ISSN 1595-093X/ E-ISSN 2315-6201)

Hassan et al/Sokoto Journal of Veterinary Sciences (2016) 14(1): 49-52 http://dx.doi.org/10.4314/sokjvs.v14i1.9

Prevalence of Newcastle disease virus antibodies in sera and eggs of helmeted guinea fowls (*Numida meleagris galeata pallas*) in Borno and Yobe States, Nigeria

SU Hassan¹*, AD El-Yuguda², HI Gambo¹, SS Baba², AG Ambali³ & IO Igbokwe¹

^{L.} Department of Veterinary Pathology, University of Maiduguri, Nigeria,

 Department of Veterinary Microbiology and Parasitology, University of Maiduguri, Nigeria, 3. Department of Veterinary Medicine, University of Maiduguri, Nigeria

*Correspondence: Tel.: +234 8038047109, E-mail: shehuhassandaudawa@yahoo.com

Abstract

The seroprevalence and maternal antibody profiles to Newcastle disease virus infection of guinea fowls were studied using haemagglutination inhibition (HI) test, in Borno and Yobe States of Nigeria. Of 822 sera and 354 egg yolk extracts tested, 327 (39.8%) and 242 (68.4%) were positive for NDV antibodies respectively. The seroprevalence was significantly higher (P< 0.05) in the dry (47%) than in the rainy (32.8%) seasons. Maternal antibodies in egg yolk extract (sampled during the rainy season), were significantly (P< 0.05) higher (68.4%) than the seroprevalence. The frequency distribution of antibody titers were skewed with 79.8% and 63.2 % of sera and egg yolk respectively having reciprocal antibody titres \leq 20. The geometric mean antibody titre was higher (P< 0.05) in the dry season (3.7±2.0) than in the rainy season (2.5±1.7). The geometric mean titre was also higher (P<0.05) in egg (8.9±2.5) than in sera (3.1±1.9). These results showed that Newcastle disease virus is enzootic among guinea fowls in Borno and Yobe states, especially in the dry season. The geometric mean titer of antibodies from egg yolk (8.9) was below the protective reciprocal titres \geq 35.4 are partially immune and may shed the virus without a clinical disease when infected thereby becoming a risk to in-contact birds.

Keywords: Egg, Guinea fowls, Maternal antibodies Newcastle disease, Seroprevalence, Received: 29-11- 2015 Acc

Accepted: 07-03-2016

Introduction

The helmeted guinea fowl is a semi- domestic to wild galiform indigenous to West Africa and is favoured for its high quality eggs and meat (Ayeni, 1983). About 6 million guinea fowls (13% of the national total) were estimated to be found in Borno and Yobe states, raised mostly on a free range scavenger system by nomadic herds men and villagers as income supplements (Hassan, 2007).

Newcastle disease (ND) is an important viral disease primarily of the chicken, as well as other species of domestic and wild birds. It is characterized by a high morbidity and mortality (Alexander *et al.*, 2012). Outbreaks of Newcastle disease in the guinea fowl have been reported in parts of the country (Haruna *et al.*, 1993), and elsewhere (Mishra *et al.*, 2001). Recently, an outbreak in guinea fowls was reported in Maiduguri, Borno state (Hassan *et al.*, 2014). Serological surveys in guinea fowls for

haemagglutination inhibition (HI) antibodies against Newcastle disease virus (NDV) have indicated an increasing prevalence of the virus around Zaria in northern Nigeria (Saidu *et al.*, 2004), and a similar trend was earlier reported in Borno and Yobe states (Ambali *et al.*, 2000). This study is a follow up to assess changes in seroprevalence and evaluate maternal antibody levels against NDV among guinea fowls in Borno and Yobe states.

Materials and Methods

Study area

The study was conducted in Maiduguri (Borno state) and Potiskum (Yobe state), in northeastern Nigeria between 9° and 13° N and 11° and 15° E. The climate is semi-arid with two seasons; a rainy season (June to October) and a dry season (November to May). The avrage annual rainfall is

650mm (Ishaku & Majid, 2010). However some areas may have as low as 500 mm (NBS, 2012).

Sample collection

A total of eight hundred and twenty-two (n=822) guinea fowl sera were collected from poultry slaughter slabs in Maiduguri and Potiskum in Borno and Yobe states respectively. These urban centers represented collection points for the surrounding countryside. Blood was collected at slaughter in plain tubes, kept slanted for 3-4 hours at room temperature (35-40 ⁰C) to clot. Subsequently the serum was harvested from each tube with a suction micropipette and stored in small plastic serum bottles in a deep freezer (Thermocool[®], Nigeria) at -20[°] C until the time of analysis. Three hundred and fifty four (n =354) fresh guinea fowl eggs were purchased from farmers in batches of 4-5 from 71 points in the rainy seasons. Antibodies were extracted from the egg yolk extract (EYE) by the method of Piela et al. (1984).

Serology

Newcastle disease virus antibodies were detected in sera and EYE by HI test using the method of Allan *et al.* (1978) as described by Hassan (2007).

Data analysis

The skewed frequency distribution of reciprocal antibody titers was normalized by log transformation after which the geometric mean titer (GMT) and standard deviation (SD) were calculated (Petrie & Watson, 2006).

GMT= antilog₁₀ {1/n ($\sum f_1 \log_{10} X_i$)}.

Where n= number tested, X_i = the reciprocal dilution and f_i = frequency.

SD = Antlog₁₀
$$\sqrt{\frac{1}{n-1}\Sigma\{f(\log_{10}X-\log_{10}X)^2\}}$$

The GMT values were compared for seasonal differences and titer differences between sera and EYE by a 2-tailed student's t- test, while seasonal differences were compared by chi-square analysis using computer software (GraphPad Instat Inc. San Diego, California 1998 version, www.GraphPadInStat.com).

Results

The prevalence of antibodies to NDV in sera and EYE of helmeted guinea fowls in Borno and Yobe states are presented in table 1. Three hundred and twenty seven (39.8%) sera were positive for HI antibodies against Newcastle disease out of the 822 tested. The seasonal distribution of positive samples showed significantly (p< 0.05) higher prevalence in the dry season (45.7%) than the rainy season (32.8%). Also, the prevalence of antibodies in EYE (68.4%) was significantly higher (p< 0.05) than that of the sera.

The GMT values for sera and eggs are presented in table 2. The frequency distribution of antibody titers were skewed to lower titers with 79.8% and

Table 1: Prevalence of antibodies against Newcastle disease virus in sera and egg yolks of helmeted guinea fowls in

 Borno and Yobe States, Nigeria

	Seasonal distrib	oution of serum antil	EYE antibody	
	Rainy	Dry	Total	titer (Rainy season)
Number tested Number (%) positive	378 124 (32.8) ^ª	444 203 (45.7) ^a	822 327 (39.8) ^b	354 242(68.4) ^b

Matched superscripts are significantly (p < 0.05) different by chi-square statistics

Table 2 : Frequencies of antibody titers in sera and egg yolks of helmeted guinea fowls and their geometric
mean titers (GMT) with standard deviations (SD)

	Frequency of antibody titer					
Reciprocal of	Seasonal distribution in serum					
antibody titer	Rainy	Dry	Total	Egg yolk		
10	61	100	161	83		
20	48	52	100	70		
40	12	39	51	44		
80	3	9	12	25		
160	0	3	3	17		
320	0	0	0	2		
640	0	0	0	1		
GMT ± SD	2.5 ± 1.7 ^ª	3.7 ± 2.0^{b}	$3.1 \pm 1.9^{\circ}$	8.9 ± 2.5 ^d		

Unmatched superscripts are significantly (p < 0.05) different by Student's t-test

63.2% having reciprocal antibody titres \leq 20 in sera and EYE respectively. Serum GMT was significantly higher in the dry (3.7 ± 2.0) than the rainy season (2.5 ± 1.7). The GMT was also significantly higher in EYE (8.9 ± 2.5) than in sera (3.1 ±1.9).

Discussion

The results of the study showed serological evidence of NDV infections in guinea fowls in Borno and Yobe states of Nigeria. The 39.8 % seroprevalence of NDV infection among guinea fowls was higher than the 23.9% reported earlier by Ambali & Aliyuda (1992). This increase supports similar trends of increasing report of seroprevalence in Zaria (Saidu et al., 2004). The enzootic nature of NDV in Nigeria, the high prevalence reported in the study area among freerange scavenger and commercial chicken flocks (Ambali et al., 2000) and the lack of vaccination and other control measures for free-range poultry including guinea fowls may explain the increasing prevalence. Guinea fowls in the study area were probably infected with NDV during their regular contact with chickens in the extensive free-range scavenger system they share. The significantly higher prevalence observed during the dry season has also been reported for free range scavenger chicken (El Yuguda et al., 2009). The spread of the disease was more extensive in the dry season possibly due to windy conditions that prevail

References

- Alexander DJ, Aldous, EW & Chad MF (2012). The long view: A selected review of 40 years of Newcastle disease research. *Avian Pathology*, **41** (4): 329-335.
- Allan WH, Lancaster JE & Toth B (1978). ND vaccines their production and use. In: *Animal Production and Health Series*. Number 10. Food and Agricultural Organization publication, Rome, Italy. Pp 1-163.
- Ambali AG & Aliyuda S (1992). Detection of haemagglutination inhibition antibodies against ND virus in sera of gray-breasted guinea fowls (*Numida meleagris galleata pallas*) in Maiduguri, Nigeria. *Annals of Borno*, **8** (9): 266-269.
- Ambali AG, Abubakar MB, Hassan SU & Adene DF (2000). Prevalence of active and passive immunity against ND in rural chickens under semi-arid conditions. *Journal of life and Environmental_Science*, **2**(3): 108-111.
- Ayeni JSO (1983). State of knowledge on the status, biology and management of grey breasted guinea-fowl (Numida meleagris galeata, Pallas). In: Helmet Guinea-fowl (Numida meleagris galeata, Pallas) in

during the hamatan, in which the virus may be transported by aerosol and dry faecal dust to distant places (Hassan, 2007).

A reciprocal titre of \geq 35.4 was reported to be protective against NDV infection (Allan *et al.*, 1978). Most of the sera (78.9%) had reciprocal titres \leq 20. This class of birds is partially immune and can succumb to a velogenic NDV infection or maintain a subclinical disease during which active excretion of the virus may occur making them a possible source of infection for in-contact birds (Parede & Young, 1990; Wambura, 2010; Kapezynski *et al.*, 2013).

Majority of the guinea fowl eggs had maternal antibodies against NDV in the EYE, with a GMT that was significantly higher than that of the sera, but was not sufficient (\geq 35.4) to protect the keets when they hatch. Outbreaks of Newcastle disease could therefore occur within the first week of life.

In conclusion, there was serological evidence of increasing, NDV infection in guinea fowls from Borno and Yobe states. The antibody titers in both sera and eggs were not protective, but could mask a subclinical infection, which may lead to virus shedding with considerable risk to in-contact birds. It is therefore recommended that guinea fowls should be protected by vaccination in the first week of life. Biosecurity should be improved for guinea fowls and other poultry on free-range management

> *Nigeria.* (TA Aire & JM Olomu, editors). Kainji Lake Research Institute pulication, New Bussa, Nigeria. Pp 10-20.

- El-Yuguda AD, Baba SS, Ibrahim UI & Brisibe F (2009). Newcastle disease and infectious bursal disease among village chickens in Borno. *Family Poultry*, **18**(1):16-23.
- Haruna ES, Shamaki AD, Echeonwu GON, Majiyagbe KA, Shuaibu Y & Du DR (1993). A natural outbreak of Newcastle disease in guinea fowl (*Numida meleagris* galeata) in Nigeria. *Revue Scientifique et Technique-Office International des Epizooties*, **12**(3): 887-893.
- Hassan SU (2007). Seroprevalence, Immunogenicity and Pathogenicity of Newcastle Disease Virus in Helmeted Guinea Fowls (Numida meleagris galleata pallas). PhD. thesis, Department of Veterinary pathology, Faculty of Veterinary Medicine, University of Maiduguri, Nigeria. Pp 1-167.
- Hassan SU, El-Yuguda AD, Gambo, HI, Maidala, HMB, Baba SS, Ambali AG & Igbokwe IO (2014). A natural outbreak of Newcastle disease among guinea fowls (*Numida meleagris galeata pallas*) in Maiduguri

North Eastern Nigeria. *Sahel Journal of Veterinary Science*, **13**(2): 63-67.

- Ishaku HT & Majid MR (2010). X-raying rainfall pattern and variability in North Eastern Nigeria: Impacts on Access to Water Supply. Journal of Water Resources and Protection, **2** (11): 952-959.
- Kapczynski DR, Afonso CL & Miller PJ (2013). Immune responses of poultry to Newcastle disease virus. *Developmental and Comparative Immunology* **41** (3): 447-453.
- Mishra S, Katrina JM, Verma, KC, Sah RL & Mishra JP (2001). Studies on the pathogenicity of Newcastle disease virus isolated in guinea fowl. *Tropical. Animal. Health and Production* **35** (5): 277-284.
- NBS (National Bureau of Statistics) (2012). Annual statistics. Published by National Bureau of Statistics, Federal Republic of Nigeria. Pp 1- 619.
- Parede L & Young PL (1990). The pathogenesis of velogenic Newcastle disease virus infection in chickens of different ages and

different levels of immunity. *Avian Dis*eases, **34** (4): 803-808.

- Petrie A & Watson P (2006). *Statistics for Veterinary and Animal Science* (Second edition). Blackwell Publishers. Oxford, UK. Pp 21-22.
- Piela TH, Gulka CM, Yates VS & Charg, PW (1984). Use of egg yolk in serological tests (ELISA and HI) to detect antibodies to Newcastle disease, infectious bronchitis and Mycoplasma gallisepticum. *Avian diseases*, **28** (4): 877-883.
- Sa'idu L, Tekdek LB & Abdu PA (2004). Prevalence of ND antibodies in domestic and semi domestic birds in Zaria, Nigeria. *Veterinarski Archiv*, **74**(4): 309-317.
- Wambura PN (2010). Detection of antibody to Newcastle disease virus in semidomesticated free-range birds (Numida meleagris and Columber livia domestica) and the risk of transmission of Newcastle disease to village chickens. Veterinarski Archiv, **80**(1):129-134.